

Multi-drug resistant facultative pathogenic bacteria colonizing the vagina of pregnant women with premature rupture of membrane, Tanzania

Emmanuel Kamgobe^{a*}, Simone Grote^{b*}, Martha F. Mushi^{c*}, Damas Wilson^a, Leticia Gandye^a, Oliver Bader^b, Stephen E. Mshana^{c*}, Uwe Groß^{b,d*}

^aDepartment of Obstetrics and Gynecology, Weill Bugando School of Medicine, Catholic University of Health and Allied Sciences Mwanza, Tanzania;

^bInstitute of Medical Microbiology, Goettingen University Medical Centre, Germany;

^cDepartment of Microbiology and Immunology, Weill Bugando School of Medicine, Catholic University of Health and Allied Sciences Mwanza, Tanzania;

^dGoettingen International Health Network

+Equal contributions

*Correspondence to Martha F. Mushi (mushimmartha@gmail.com)

ABSTRACT

Background: Premature rupture of membrane (PROM) contributes to approximately one-third of premature birth and 10% perinatal mortality worldwide. Here, we report the patterns of facultative pathogenic bacteria colonizing the vagina of pregnant women to guide prophylactic antibiotic treatment in the management of PROM.

Methods: This comparative cross-sectional study was conducted between August 2015 and March 2016. High vaginal swabs were collected and processed to detect the presence of facultative pathogenic bacteria. Isolate identification and antibiotic susceptibility testing was conducted using MALDI-TOF MS and VITEK-2 system, respectively. Data were analyzed using STATA version 13.

Results: A total of 175 pregnant women with PROM and 175 without PROM were investigated. The median age of the pregnant women with PROM was significantly higher than that of pregnant women without PROM: 27 [21-32] vs. 25 [21-29], $p=0.026$. Pregnant women with PROM were significantly more likely to be colonized with facultative pathogenic bacteria 59/175 (33.7%), 95% CI: 26.7-40.7 than pregnant women without PROM; 27/175 (15.4%), 95% CI: 10.1-20.7, $P<0.001$. *Escherichia coli* were significantly more isolated from pregnant women with PROM than those without PROM: 36 (73.5%) vs. 13 (26.5%), $p<0.001$. The proportion of resistance among pathogenic isolates from women with PROM to ampicillin, trimethoprim/sulfamethoxazole and cefotaxime were 100%, 66.7% and 40%, respectively.

Conclusions: The vagina of pregnant women with PROM is significantly more colonized by multi-resistant facultative pathogenic bacteria than that of pregnant women without PROM. Further studies should be done to elucidate the impact of these bacteria in relation to PROM and the pregnancy outcome.

Key words: Vaginal colonization, facultative pathogenic bacteria, pregnancy, premature rupture of membrane, multi drug resistance

INTRODUCTION

Worldwide, premature rupture of membranes (PROM) among pregnant women has been found to range from 3.3% to 10%, with 80% of them occurring at term^{1,2}. PROM leads to the loss of the natural protection of the fetus hence posing a threat to bacterial

infections³. In addition, PROM is highly associated with the increased pregnancy complications such as preterm labor, fetal demise, respiratory distress syndrome, neonatal sepsis, umbilical cord prolapse, postpartum endometritis, disseminated intravascular coagulopathy (DIC) and chorioamnionitis^{1,4}. The pathogenesis of PROM has been linked to the isolation of facultative

pathogenic bacteria in the vagina⁵⁻⁸, there is a strong association between pathogens colonizing the vagina and subsequent chorioamnionitis. Bacteria and protozoan parasites (*Trichomonas vaginalis*) secrete proteases and other factors that degrade the collagen and weaken the fetal membrane^{9,10}. Furthermore, host inflammatory responses due to pathogenic bacteria can induce the production of prostaglandin which can lead to uterine irritability and membrane collagen degradation hence increasing the risk of PROM^{8,11}.

In developing countries, *Escherichia coli*, *Klebsiella pneumoniae*, Group B Streptococcus (GBS), *Staphylococcus aureus* and *Streptococcus pyogenes* have been found to be the commonest facultative pathogenic bacteria colonizing the vagina and implicated in PROM^{5-7,12}. These pathogenic bacteria have also been implicated in chorioamnionitis^{13,14}. Despite 12% contribution of the PROM to antenatal hospital admissions in East Africa¹⁵ and black women being reported to have the highest risk of being colonized by the potential pathogenic bacteria¹⁶, the spectrum of the respective bacteria colonizing the vagina of pregnant women with and without PROM in East Africa is not well understood.

A previous study¹⁷ in Uganda among pregnant women with PROM, noted the resistance to most commonly used antibiotics such as ceftriaxone, ampicillin, trimethoprim/sulfamethoxazole and erythromycin was high with good susceptibility to expensive antibiotics such as vancomycin and meropenem. The World Health Organization (WHO) recommends the use of antibiotic prophylaxis among pregnant women with PROM¹⁸. Different regimen involving penicillin and erythromycin have been recommended. However, number of factors should be assessed such as population of women to be offered antenatal prophylactic antibiotics and types of antibiotics to be used. With increase antibiotic resistance among the pathogens involved in PROM local susceptibility data are crucial in establish empiric treatment protocol with emphasis on the individual tailored treatment. This comparative cross-sectional study was designed to investigate the patterns of facultative pathogenic bacteria colonizing pregnant women with and without PROM and their antimicrobial susceptibility.

METHODS

Study design, area, and population

The comparative cross-sectional study was conducted between August 2015 and March 2016 at Bugando Medical Centre (BMC), Sekou Toure Regional Hospital, Nyamagana District Hospital and Buzuruga Health Center in the city of Mwanza, Tanzania. The selected hospital are

the highly populated faith base and government hospital which cover large percentage of the Mwanza population. Bugando Medical Centre is the tertiary consultant teaching hospital of Catholic University of Health and Allied Sciences with bed capacity of 900. Sekou Toure Regional Referral hospital is located in Ilemela district with bed capacity of 375 serving the all referrals from six district hospitals of Mwanza region. While Nyamagana District hospital has maternity bed capacity of 30 and Buzuruga health center has maternity ward with a bed capacity of 15. The study involved all pregnant women with gestation age of 36 weeks and above with and without PROM during the study period.

Sampling, inclusion, and exclusion criteria

Using Kirkwood formula for comparative studies¹⁹ and assumption of the 10% effect size by the prevalence obtained in the previous study¹², the minimum sample size obtained was 350 pregnant women (175 pregnant women with and 175 pregnant women without PROM). The pregnant women with and without PROM were recruited serially until the sample size was reached. The study excluded all pregnant women with cervical incompetency, polyhydramnios, mal-presentation, multiple pregnancies, fever, abdominal pain, foul smelling per vaginal leakage and history of antibiotic therapy in the past two weeks prior to the study to minimize the bias in relation to the colonization of the facultative bacteria. In the current study, pathogenic bacteria were defined as bacteria which are potentially capable of causing clinical infections in the genital urinary tract²⁰.

Sample collection and laboratory procedures:

By the use of sterile Cusco speculum, cervix was exposed, and high vaginal swab was taken using a sterile swab. The swabs were transported to the microbiology laboratory using Stuart transport media (HiMedia, India) within 2 hours of collection. All swabs were cultured on the 5% sheep blood agar (BA) and MacConkey agar (MCA) (Oxoid, UK) and aerobically incubated at 37°C for 24-48 hours. Identification to species level was done by matrix-assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry (Bruker Daltonics, Germany) on extracted cells as previous described^{21,22}. Only facultative pathogenic bacteria were included for statistical analysis and subsequent antimicrobial susceptibility testing. Potential contaminants, e.g. *S. epidermidis*, *Bacillus* spp., were excluded. The current study mainly concentrated on the antibiotics that are recommended for PROM management. The tested antibiotics included: ampicillin (AMP), trimethoprim/sulfamethoxazole, (SXT), ciprofloxacin (CIP), Gentamicin (CN), ceftazidime (CAZ), ceftriaxone

(CRO) and meropenem (MRP) (Oxoid, UK). Antimicrobial susceptibility testing was done using VITEK-2 system (bioMérieux, France) and interpreted as per EUCAST (http://www.eucast.org/clinical_breakpoints/) guidelines. Using excel sheet, data were double entered, cleaned and transferred to STATA version 11 for analysis. Categorical variables such as residence, marital status, education, occupation, gravidity, history of PROM, number of antenatal care (ANC) visit, presence or absence of PROM and positive or negative bacterial growth were summarized as proportions. Continuous data (age, gestation age and parity) were summarized using median and inter quartile range. The statistical significance was set at a p value of less than 0.05. Two-sample test of proportions was used to compare the pattern of facultative pathogenic bacteria colonization among pregnant women with PROM and those without PROM.

Ethics approval and consent to participate

The joint CUHAS/BMC research ethics and review committee granted ethical clearance with certificate number CREC/096/2015. Permission to conduct the study was sought from all hospital administrations. All patients were requested to sign a written informed consent before recruitment was done.

RESULTS

Demographic characteristic

A total of 350 pregnant women (175 with PROM and 175 without PROM) were enrolled and analyzed. The majority of studied women resided in urban areas 286(81.7%) and had primary school education 185 (52.9%). The median age of pregnant women was 26 [21-31] years. The median age of pregnant women with PROM was significantly higher than pregnant women without PROM 27 [21-32] vs. 25 [21-29], $p=0.026$ (**Table 1**).

The median gestation age of pregnant women at the time of enrollment was 38[36-40] weeks for pregnant women with PROM and 38[37-39] for pregnant women without PROM. History of PROM in previous pregnancy was

higher among pregnant women with PROM than pregnant women without PROM 20 (11.4%) vs. 6 (3.4%), $p=0.004$. In addition, history of preterm birth in the previous pregnancy was higher among pregnant women with PROM than those without PROM 16(72.7%) vs. 6 (27.3%), $p=0.026$. Regarding antenatal visits, high proportion of pregnant women with PROM had more than 4 antenatal visits compared to pregnant women without PROM 92 (52.6%) vs. 54 (30.9%), $p<0.001$ (**Table 1**).

Bacterial colonization pattern

Of 350 women screened, 86 (24.6%) were colonized with facultative pathogenic bacteria. Pregnant women with PROM were significantly more often colonized with facultative pathogenic bacteria 59/175 (33.7%), 95% CI; 26.7-40.7 than pregnant women without PROM 27/175 (15.4%), 95% CI; 10.1-20.7, $p<0.001$. The most frequently isolated bacteria were *Escherichia coli* 49(39.2%) and *Pseudomonas spp.* 22(17.6%). *E. coli* isolates were significantly more from pregnant women with PROM than those without PROM, 36 (73.5%) vs. 13 (26.5%), $p<0.001$ (**Table 2**). Of 175 pregnant women with PROM, 13(7.4%) had double colonization with two different species of facultative pathogenic bacteria. All pathogenic bacteria isolated from pregnant women with PROM were resistant to ampicillin, while those isolated from pregnant women without PROM were 92.6% (25/26) resistant to ampicillin. The proportion of resistant bacteria from women with PROM to ampicillin, trimethoprim/sulfamethoxazole and cefotaxime were 57(100%), 46(66.7%) and 28(40%), respectively while for women without PROM the proportion was 25(96.2%), 21(52.5%) and 20(48.8%), respectively **Table 3**. *E. coli* isolates from pregnant women with PROM were significantly more resistant to trimethoprim/sulphamethoxazole than *E. coli* isolates from women without PROM 82.9% (29/35) vs. 58.3% (7/12), $p<0.001$ **Figure 1**. One *E. coli* isolate from pregnant woman with PROM was resistant to all antibacterial agents tested (ampicillin, ciprofloxacin, gentamycin, trimethoprim/sulphamethoxazole, ceftriaxone, ceftazidime, cefotaxime, ertapenem, meropenem).

TABLE 1. Socio-demographic and clinical data of 350 pregnant women studied

Patient characteristic	Total n=350 (%)	PROM n=175 (%)	Non-PROM n=175 (%)	p-value
Age* years	26 [21-31]	27 [21-32]	25 [21-29]	0.026
Residence				
Urban	286 (81.7)	131 (74.86)	155 (88.6)	
Rural	64 (18.3)	44 (25.14)	20 (11.4)	0.001
Marital status				
Single	73 (20.85)	46 (26.3)	27 (15.44)	
Married	277(79.15)	129 (73.7)	148 (84.57)	0.013
Education				
Primary	185(52.85)	91 (52)	89 (50.9)	
Secondary	123(35.14)	68 (38.9)	55 (31.4)	
University	42 (12)	16 (9.14)	31 (17.7)	0.071
Gravidity				
Prime	100(28.6)	36 (20.6)	64 (36.6)	
Gravid 2	175(50.0)	86 (49.0)	89 (50.9)	
Multigravida	75 (21.4)	53 (30.3)	22 (12.6)	<0.001
GA* weeks	38 [36-40]	38 [36-40]	38 [37-39]	0.012
PROM before				
No	324(92.6)	155 (88.6)	169 (96.6)	
Yes	26 (7.4)	20 (11.4)	6 (3.4)	0.004
GA at booking*	20 [18-22]	20 [18-22]	20 [18-22]	0.236
ANC visit				
Below 4	79 (22.6)	29 (16.6)	50 (28.6)	
4	125(35.7)	54 (30.9)	71 (40.6)	
Above 4	146(41.7)	92 (52.6)	54 (30.9)	<0.001

GA is gestation age, * Are variables were median is the measure of central tendency

TABLE 2. Vaginal pathogenic bacteria colonizing 350 pregnant women in Mwanza

BACTERIA	PROM N (%)	Non-PROM N (%)	p- value
<i>E. coli</i> (49)	36 (73.5)	13 (26.5)	<0.001
<i>Pseudomonas spp.</i> (22)	12 (54.6)	10 (45.5)	0.273
<i>K. pneumoniae</i> (17)	11 (64.7)	6 (35.3)	0.043
<i>Enterobacter spp.</i> (13)	8(61.5)	5(38.5)	0.119
Pathogenic GPB* (10)	9(90)	1(10)	0.002
<i>Acinetobacter spp.</i> (7)	1(14.3)	6(85.7)	0.0038
Other GNB* (7)	5(71.4)	2(28.6)	0.054
Total (125)	82(65.6)	40(34.4)	<0.001

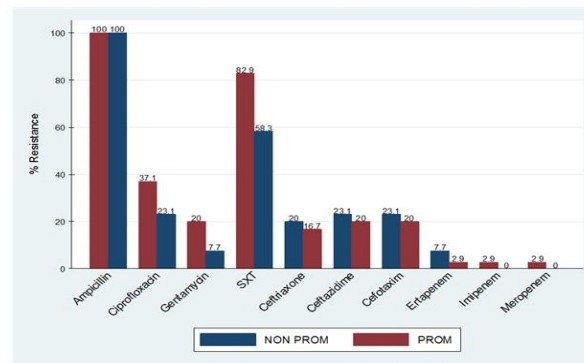
*Pathogenic GPB stands for pathogenic gram-positive bacteria which include *S. aureus*, *S. haemolyticus*, *E. faecalis* and *S. saprophyticus* and other GNB stands for other gram-negative bacteria which include *Proteus spp.*, *Morganella morganii* and *Escherichia hermannii*.

TABLE 3. Antimicrobial resistance pattern among vagina pathogenic bacteria isolates

Antibiotic	PROM		Non-PROM	
	Tested	Resistant (%)	Tested	Resistant (%)
Ampicillin	57	57 (100)	26	25 (96.2)
Ciprofloxacin	69	22 (31.9)	41	7 (17.1)
Gentamycin	69	16 (23.2)	41	8 (19.5)
SXT*	69	46 (66.7)	40	21 (52.5)
Ceftriaxone	57	14 (24.6)	22	3 (13.6)
Ceftazidime	70	17 (24.3)	40	9 (22.5)
Cefotaxim	70	28(40)	41	20 (48.8)
Ertapenem	57	1(1.8)	26	1 (4)
Imipenem	68	3(4.4)	41	0 (0.0)
Meropenem	68	2(2.9)	41	0 (0.0)

*SXT: trimethoprim/sulphamethaxazole

FIGURE 1: Antibacterial resistance pattern of E. coli colonizing vagina of pregnant women with and without premature rupture of membrane



*SXT: trimethoprim/sulphamethaxazole

DISCUSSION

Premature rupture of the membrane (PROM) contributes to approximately one third of premature births^{2,23,24}, and approximately 10% of perinatal mortality²⁵. A significant proportion of pregnant women with PROM in the current study had history of PROM and premature delivery in the previous pregnancy. This has also been reported elsewhere^{26, 27} and could be due to the possibility of the pregnant woman genetic defect in collagen synthesis that can affect the structure and function of the fibrillar collagens²⁸. The weakening of the connective tissue by the enzymatic depolarization of the collagen fibers in fetal membrane can also explain the observed findings^{29,30}. Women with PROM in the current study were significantly older than those without PROM pointing to the possibility of age-dependent collagen synthesis³¹. However, there is no documentation of the influence of age and facultative pathogenic bacteria colonizing vagina during the reproductive age. The age has been found to influence normal flora before puberty and after menopause^{32, 33}.

Vaginal colonization with facultative pathogenic bacteria was significantly more often observed in pregnant women with PROM than in pregnant women without PROM. This has also been reported in previous studies in Tanzania and India^{12,25}. Bacteria in the vagina have been found to secrete enzymes that can either degrade the fetal membranes or increase production of prostaglandins^{4,12,34-39} high concentration of prostaglandins can stimulate the uterine contractions leading to the membrane rupture. This is further supported by the fact that the presence of pathogenic bacteria has been associated with chorion thinning among PROM pregnant women⁴⁰.

As it was also previously reported in Tanzania and India^{12,41}, *E. coli* and *Pseudomonas* spp. were the commonest bacteria detected. This could partly be explained by the fact that these pathogens belong to the normal flora of the gastrointestinal tract and might therefore be present in perineum with increased chance to colonize the genital tract. *Staphylococcus aureus* was the commonest gram-positive bacterial species detected among pregnant women with PROM in this study. Similar observations were made previously^{12, 25}. Detection of *S. aureus* colonizing women with PROM has been linked with other factors like urinary tract infections and bacterial vaginosis²⁵, these factors were not investigated in the current study.

Bacteria isolated from pregnant women with PROM were more resistant to ampicillin and trimethoprim/sulfamethoxazole. Similar results have been reported in other studies from pregnant women^{42,43} and post-delivery women in similar settings⁴⁴. This could partly be explained by the fact that ampicillin and

trimethoprim/sulfamethoxazole are the commonest class of antibiotic in use in the study settings. Despite the fact that this study has clearly demonstrated the significant differences in patterns of bacteria colonizing pregnant women with PROM and those without PROM, neonatal outcomes which could give more evidence on the association of vagina colonization of the facultative pathogenic bacteria and clinical fetal infections especially for the pregnant women with PROM were not recorded. Furthermore, the presence of *Candida* species were not assessed, this has been recommended for future studies.

In conclusion, vagina of pregnant women with PROM was more colonized by multi-resistant facultative pathogenic bacteria than the one of pregnant women without PROM. *Escherichia coli* strains were the commonest pathogenic bacteria and were highly resistant to ampicillin and trimethoprim/sulphamethoxazole. Further studies should be done to elucidate the impact of these pathogens in relation to PROM and the pregnancy outcome. There is a need to adjust the empirical prophylaxis treatment of PROM based on the local susceptibility profile.

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REFERENCES

1. Tavassoli F, Ghasemi M, Mohamadzade A, F T, S J, editors. (Survey of Pregnancy Outcome in Preterm Premature Rupture of Membranes with Amniotic Fluid Index < 5 and ≥ 5). onjournal; 2010: Mashhad university of medical sciences.
2. Mercer BM, Miodovnik M, Thurnau GR, Goldenberg RL, Das AF, Ramsey RD, et al. Antibiotic therapy for reduction of infant morbidity after preterm premature rupture of the membranes: a randomized controlled trial. *Jama*. 1997;278(12):989-95.
3. Poma PA. Premature rupture of membranes. *Journal of the National Medical Association*. 1996;88(1):27.
4. Donati L, Di Vico A, Nucci M, Quagliozzi L, Spagnuolo T, Labianca A, et al. Vaginal microbial flora and outcome of pregnancy. *Archives of gynecology and obstetrics*. 2010;281(4):589-600.
5. McDonald HM, O'Loughlin JA, Jolley PT, Vigneswaran R, McDonald PJ. Changes in vaginal flora during pregnancy and association with preterm birth. *Journal of Infectious Diseases*. 1994;170(3):724-8.
6. Regan J, Chao S, James L. Premature rupture of membranes, preterm delivery, and group B streptococcal colonization of mothers. *American journal of obstetrics and gynecology*. 1981;141(2):184-6.
7. EKWO EE, GOSSELINK CA, WOOLSON R, MOAWAD A. Risks for premature rupture of amniotic membranes. *International Journal of Epidemiology*. 1993;22(3):495-503.
8. McGregor JA, French JI, Parker R, Draper D, Patterson E, Jones W, et al. Prevention of premature birth by screening and treatment for common genital tract infections: results of a prospective controlled evaluation. *American journal of obstetrics and gynecology*. 1995;173(1):157-67.
9. MCGREGOR JA, FRENCH JI, TODD JK, LAWELLIN D, FRANCO-BUFF A, SMITH C. Bacterial Protease-Induced Chorionic Membrane Reduction of Strength and Elasticity. *Obstetrics & Gynecology*. 1987;69(2):167-74.

10. Draper D, Jones W, Heine RP, Beutz M, French JI, McGregor JA. *Trichomonas vaginalis* weakens human amniochorion in an in vitro model of premature membrane rupture. *Infectious diseases in obstetrics and gynecology*. 1995;2(6):267-74.
11. McGregor J, Schoonmaker J, Lunt B, Lawellin D. Antibiotic inhibition of bacterially induced fetal membrane weakening. *Obstetrics & Gynecology*. 1990;76(1):124-8.
12. August F. The microbial pattern associated with preterm premature rupture of membranes as seen at Muhimbili National Hospital: Muhimbili University of Health and Allied Sciences; 2007.
13. Sherman D, Tovbin J, Lazarovich T, Avrech O, Reif R, Hoffmann S, et al. Chorioamnionitis caused by gram-negative bacteria as an etiologic factor in preterm birth. *European Journal of Clinical Microbiology and Infectious Diseases*. 1997;16(6):417-23.
14. Martius J, Eschenbach D. The role of bacterial vaginosis as a cause of amniotic fluid infection, chorioamnionitis and prematurity—a review. *Archives of gynecology and obstetrics*. 1990;247(1):1-13.
15. Nakubulwa S, Kaye DK, Bwanga F, Tumwesigye NM, Mirembe FM. Genital infections and risk of premature rupture of membranes in Mulago Hospital, Uganda: a case control study. *BMC research notes*. 2015;8(1):573.
16. Goldenberg RL, Klebanoff MA, Nugent R, Krohn MA, Hillier S, Andrews WW. Bacterial colonization of the vagina during pregnancy in four ethnic groups. *American Journal of Obstetrics & Gynecology*. 1996;174(5):1618-21.
17. Musaba MW, Kagawa MN, Kiggundu C, Kiondo P, Wandabwa J. Cervicovaginal bacteriology and antibiotic sensitivity patterns among women with premature rupture of membranes in Mulago Hospital, Kampala, Uganda: A cross-sectional study. *Infectious diseases in obstetrics and gynecology*. 2017;2017.
18. WHO Recommendation on the prophylactic antibiotic of choice in women with preterm prelabour rupture of membranes. The WHO reproductive Health Library November 2015.
19. Kirkwood B, Sterne J. Calculation of required sample size. London: Blackwells Science Limited. 1988.
20. Ronald AR, Alfa MJ. *Microbiology of the genitourinary system*. 1996.
21. Wieser A, Schneider L, Jung J, Schubert S. MALDI-TOF MS in microbiological diagnostics—identification of microorganisms and beyond (mini review). *Applied microbiology and biotechnology*. 2012;93(3):965-74.
22. Bader O, Weig M, Taverne-Ghadwal L, Lugert R, Gross U, Kuhns M. Improved clinical laboratory identification of human pathogenic yeasts by matrix-assisted laser desorption ionization time-of-flight mass spectrometry. *Clinical Microbiology and Infection*. 2011;17(9):1359-65.
23. Kaya D. Risk factors of preterm premature rupture of membranes at Mulago hospital Kampala. *East African medical journal*. 2001;78(2):65-9.
24. Eleje GU, Adinma JI, Ghasi S, Ikechebelu JI, Igwegbe AO, Okonkwo JE, et al. Antibiotic susceptibility pattern of genital tract bacteria in pregnant women with preterm premature rupture of membranes in a resource-limited setting. *International Journal of Gynecology & Obstetrics*. 2014;127(1):10-4.
25. Karat C, Madhivanan P, Krupp K, Poornima S, Jayanthi N, Suguna J, et al. The clinical and microbiological correlates of premature rupture of membranes. *Indian journal of medical microbiology*. 2006;24(4):283.
26. Doody D, Patterson M, Voigt L, Mueller B. Risk factors for the recurrence of premature rupture of the membranes. *Paediatric and perinatal epidemiology*. 1997;11(S1):96-106.
27. Ladfors L, Mattsson L-Å, Eriksson M, Milsom I. Prevalence and risk factors for prelabor rupture of the membranes (PROM) at or near term in an urban Swedish population. *Journal of perinatal medicine*. 2000;28(6):491-6.
28. Anum EA, Hill LD, Pandya A, Strauss J. Connective tissue and related disorders and preterm birth: clues to genes contributing to prematurity. *Placenta*. 2009;30(3):207-15.
29. Bourne G. The foetal membranes: a review of the anatomy of normal amnion and chorion and some aspects of their function. *Postgraduate medical journal*. 1962;38(438):193.
30. Strauss JF. Extracellular matrix dynamics and fetal membrane rupture. *Reproductive Sciences*. 2013;20(2):140-53.
31. Varani J, Dame MK, Rittie L, Fligel SE, Kang S, Fisher GJ, et al. Decreased collagen production in chronologically aged skin: roles of age-dependent alteration in fibroblast function and defective mechanical stimulation. *The American journal of pathology*. 2006;168(6):1861-8.
32. Cauci S, Driussi S, De Santo D, Penacchioni P, Iannicelli T, Lanzafame P, et al. Prevalence of bacterial vaginosis and vaginal flora changes in peri- and postmenopausal women. *Journal of clinical microbiology*. 2002;40(6):2147-52.
33. Burton JP, Reid G. Evaluation of the bacterial vaginal flora of 20 postmenopausal women by direct (Nugent score) and molecular (polymerase chain reaction and denaturing gradient gel electrophoresis) techniques. *Journal of Infectious Diseases*. 2002;186(12):1770-80.
34. Yudin MH, Money DM. Screening and management of bacterial vaginosis in pregnancy. *Journal of obstetrics and gynaecology Canada: JOGC= Journal d'obstetrique et gynecologie du Canada: JOGC*. 2008;30(8):702-16.
35. Jones F, Miller G, Gadea N, Meza R, Leon S, Perez J, et al. Prevalence of bacterial vaginosis among young women in low-income populations of coastal Peru. *International journal of STD & AIDS*. 2007;18(3):188-92.
36. Baisley K, Chagalucha J, Weiss HA, Mugeye K, Everett D, Hambleton I, et al. Bacterial vaginosis in female facility workers in north-western Tanzania: prevalence and risk factors. *Sexually transmitted infections*. 2009;85(5):370-5.
37. Shayo PA, Kihunrwa A, Massinde AN, Mirambo M, Rumanyika RN, Ngwalida N, et al. Prevalence of bacterial vaginosis and associated factors among pregnant women attending at Bugando Medical Centre, Mwanza, Tanzania. *Tanzania journal of health research*. 2012;14(3).
38. Epstein FH, Parry S, Strauss JF. Premature rupture of the fetal membranes. *New England Journal of Medicine*. 1998;338(10):663-70.
39. Novy MJ, McGregor JA, Iams JD. New perspectives on the prevention of extreme prematurity. *Clinical obstetrics and gynecology*. 1995;38(4):790-808.
40. Fortner B, Grotegut CA, Ransom CE, Bentley RC, Feng L, Lan L, et al. Bacteria localization and chorion thinning among preterm premature rupture of membranes. *PLoS one*. 2014;9(1):e83338.
41. Kerur BM, Bhat BV, Harish B, Habeebullah S, Kumar CU. Maternal genital bacteria and surface colonization in early neonatal sepsis. *The Indian Journal of Pediatrics*. 2006;73(1):29-32.
42. Masinde A, Gumodoka B, Kilonzo A, Mshana S. Prevalence of urinary tract infection among pregnant women at Bugando Medical Centre, Mwanza, Tanzania. *Tanzania journal of health research*. 2009;11(3).
43. Chaula T, Seni J, Ng'walida N, Kajura A, Mirambo MM, DeVinney R, et al. Urinary Tract Infections among HIV-Positive Pregnant Women in Mwanza City, Tanzania, Are High and Predicted by Low CD4. *International journal of microbiology*. 2017;2017.
44. Nelson E, Kayega J, Seni J, Mushi MF, Kidenya BR, Hikororo A, et al. Evaluation of existence and transmission of extended spectrum beta lactamase producing bacteria from post-delivery women to neonates at Bugando Medical Center, Mwanza-Tanzania. *BMC research notes*. 2014;7(1):1.

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